

# The changes of inflammatory markers and irisine level in the first year of insulin therapy in type 2 diabetes patients

<sup>1</sup>Cristian-Ioan Crăciun, <sup>2,3</sup>Anca-Elena Crăciun, <sup>1</sup>Ștefan C. Vesa, <sup>1</sup>Raluca M. Pop, <sup>1</sup>Corina I. Bocșan, <sup>1</sup>Anca D. Buzoianu

<sup>1</sup> Department of Pharmacology, Toxicology and Clinical Pharmacology, University of Medicine and Pharmacy “Iuliu Hatieganu” Cluj-Napoca, Romania; <sup>2</sup> Department of Diabetes, Nutrition and Metabolic Diseases, University of Medicine and Pharmacy “Iuliu Hatieganu” Cluj-Napoca, Romania; <sup>3</sup> „Regina Maria” Policlinic, Cluj-Napoca, Romania.

**Abstract.** Objective: To evaluate the changes of inflammatory markers and irisine plasma level in the first year after initiation of insulin therapy in type 2 diabetes patients (T2DM). Material and Method: Twenty-three T2DM patients who started insulin therapy for better glycemic control were enrolled in our study. The study was conducted during the first year of insulin therapy and included 4 visits: initial visit, after 3 months, after 6 months, and after one year of insulin-therapy. Anthropometric and metabolic parameters (basal glycaemia, HbA1c, lipid profile) were measured at each visit, while adiponectin, interleukin-1 $\beta$  and irisin at the beginning, after 6 months and after one year of insulin therapy. Results: The mean age of the 23 patients was  $62.30 \pm 10.29$  years, with a median duration of T2DM of 7 years (3-12 years). During the first year of insulin therapy the patients presented significant modification of HbA1c ( $10.23 \pm 1.94\%$  versus  $7.63 \pm 0.78\%$ ,  $p < 0.001$ ) and basal glycaemia ( $248.17 \pm 61.84$  mg/dl versus  $151.89 \pm 41.72$  mg/dl,  $p < 0.001$ ), with significant increase in body weight and skeletal muscle mass, and no significant increase in body fat mass or percent of body fat. Adiponectin level was significantly lower after first year of insulin therapy:  $7.06$  mg/L (3.98;10.01) versus  $4.81$  mg/L (3.02;6.95),  $p = 0.044$ ; irisin level was significantly higher after one year of insulin therapy:  $0.65$  ng/ml (0.00;1.63) versus  $1.37$  ng/ml (0.57;1.70),  $p = 0.004$ , with no significant change in circulating IL-1 $\beta$  levels:  $5.45$  pg/ml (3.54;7.22) versus  $5.15$  pg/ml (3.65;6.70),  $p = 0.076$ . Conclusion One year after insulin therapy initiation, patients with T2DM showed a significant decrease in adiponectin levels, a significant increase in irisin levels and no significant changes in IL-1 $\beta$  levels.

**Key Words:** type 2 diabetes mellitus, insulin therapy, adiponectin, interleukin - 1 $\beta$ , irisin.

**Copyright:** This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Corresponding Author:** A. E. Craciun, email: [anca.craciun@umfcluj.ro](mailto:anca.craciun@umfcluj.ro).

## Introduction

Despite multifactorial clinical management of the patient with type 2 diabetes mellitus (T2DM), nearly two-thirds of them die due to cardiovascular (CV) events (Herman et al 2017). The use of insulin therapy in T2DM patients was proved to be useful for significant reduction of microvascular complications in landmark randomized clinical trials such as UKPDS (United Kingdom Prospective Diabetes Study) due to an intensive and sustained glycemic control (UKPDS Group 1998), even though initial data showed no reduction in macrovascular risk. The follow-up of these patients has shown that the initial glycemic control offered further protection for microvascular complications, while subsequently published data showed that intensive glycemic control at the onset of T2DM had long-term benefits regarding macrovascular complications (Holman et al 2008). But in real-life management of T2DM, the majority of patients receiving insulin therapy are those with older duration of the disease and important comorbidities, which are by themselves risk factors for adverse effects (Ferrannini & DeFronzo 2015). In ORIGIN (Outcome Reduction with an Initial Glargine Intervention) trial, the use of insulin in T2DM patients with

cardiovascular disease (CVD) or CV risk factors showed no safety concerns regarding mortality or CVD events (Gerstein et al 2012). On the other hand, Gamble et al observed a significant and graded association between level of exposure to insulin and CV and non-vascular mortality (Gamble et al 2010). These findings were also supported by the observations from ACCORD study (which was stopped due to increased mortality in the arm with intensive glucose control) (Riddle et al 2010) and by further analysis of Bonds et al, which showed a proportionally increased risk for higher insulin dose (Bonds et al 2010). Therefore, the published results about the effect of insulin on CVD and mortality have contradictory results and methodological limitations, no final conclusion being provided regarding risk-benefit of insulin therapy in T2DM. (Muis et al 2005; Asleh et al 2017). Beyond antihyperglycaemic action, insulin has pleiotropic effects. In vitro studies proved that insulin has anti-inflammatory and anti-atherogenic effect (Dandona P et al 2009). Its action on vascular endothelium leads to nitric-oxide release and vasodilatation (Grover A et al 1995) and has a suppressive effect on the expression of NF $\kappa$ B and MCP-1 in human endothelial cells (Aljada et al 2001).

Adiponectin is a major cytokine secreted by adipose tissue, which has been shown in fundamental studies to play an important role in glucose metabolism by reducing insulin resistance and inflammation (Kadowaki *et al* 2006). High plasma concentration of adiponectin was linked with lower risk of T2DM (Nicholson *et al* 2018; Yamamoto *et al* 2014) and its serum level was inversely correlated with weight and central obesity in humans, regardless of diabetic status (Nayak *et al* 2010). In patients with T2DM, adiponectin level is lower than in non-diabetic controls and its levels were negatively correlated with insulin resistance and adiposity (Aleidi *et al* 2015; Nayak *et al* 2010). There is a crosstalk between adiponectin and insulin: the adipokine has an insulin-sensitizing action, due to muscle microvasculature dilatation, with increasing delivery of insulin to muscle cells (Zhao *et al* 2014), but there are little evidence regarding the effect of insulin therapy on adiponectin levels in T2DM.

Irisin is a newly discovered myokine, whose secretion is influenced by (PGC1) - $\alpha$  (peroxisome proliferator-activated receptor- $\alpha$  coactivator-1) (Huh *et al* 2012). Irisin has been proposed as the mediator of beneficial effects of physical exercise on metabolism by increasing thermogenesis due to a browning process of the white fat (Boström *et al* 2012). The response of genes encoding irisin after exercise is different depending on age, higher increase in the elderly than in young, but lower circulating levels of irisin due to reduced muscle mass in the elderly (Timmons *et al* 2012). Thus, the circulating level of irisin is inversely proportional to the circulating level of adiponectin and direct proportionally with body mass index (BMI), basal glycaemia and total cholesterol (Enerback 2010).

Since contradictory literature data regarding the response of inflammatory markers in patients with T2DM treated with insulin is reported, and since so far, few data of insulin therapy effect on irisin levels were reported, our aim was to conduct a personal research in order to assess the impact of insulin therapy initiation on inflammatory markers, adiponectin and on irisin plasma level in T2DM patients.

## Material and method

We conducted a cohort study that included patients with prior diagnosis of T2DM who needed initiation of insulin therapy for better glycemic control. All patients received basal insulin therapy with glargine (Lantus, Sanofi). The study was conducted during the first year of insulin therapy and included 4 visits: initial visit, after 3 months, after 6 months, and after one year of insulin therapy. The study was approved by the Ethics Committee of the "Iuliu Hațieganu" University of Medicine and Pharmacy, Cluj-Napoca.

At the initial visit, the patients received informations about the study and the Informed Consent was signed. We collected information about patients' data: age, gender, duration of diabetes, current treatment, complications, blood tests (glycaemia, HbA1c, creatinine, transaminase, LDL cholesterol, HDL cholesterol, triglycerides, irisine, adiponectin, interleukin 1-beta). During this visit patients received information about lifestyle optimization and were trained in insulin injection and blood glucose monitoring technic, received information about insulin dose adjustment algorithms and about hypoglycemia management. Afterwards, during subsequent visits, the procedures for the initial clinical and paraclinical measurements were performed. Adiponectin,

IL-1 $\beta$  and irisin levels were measured at initial visit, after 6 months and after 12 month after the initiation of insulin therapy. The body composition analysis was performed during every visit, using InBody 720 device (Biospace Co., Korea) according to the recommendation provided in the user manual (InBody 720 User's Manual, Biospace 1996-2004). This analysis was derived from the four compartment model, which divides body composition into four components: total body water, protein, mineral and body fat mass.

After prelevation, blood samples (5 mL) were collected using pyrogen-free tubes without anticoagulant. Samples were immediately centrifuged at 3000 g, for 10 min at 4 °C. Serum was separated in Eppendorf tubes and frozen at -80 °C until analysis. The serum levels of irisin, adiponectin and IL 1 (Abbexa USA kits) were measured using the enzyme-linked immunosorbent assay ELISA using commercially available reagents. The standard dilutions and samples preparation was performed according to the manufacturer's instructions.

Statistical analysis was performed using MedCalc Statistical Software version 18.2.1 (MedCalc Software bvba, Ostend, Belgium; <http://www.medcalc.org>; 2018). Quantitative data were tested for normality of distribution using the Kolmogorov-Smirnov test, and were expressed by mean and standard deviation or median and 25-75 percentiles. Comparison between measurements was performed using ANOVA for repeated measures or Friedman test, whenever appropriate. A p value <0.05 was considered statistically significant.

## Results

Twenty-five patients were enrolled in the study, of which 23 patients reached the final visit (7 women, 30.4%), while two were excluded from the study (the first one was excluded because he did not attend all protocol visits while the second patient discontinued insulin therapy before the final visit). The mean age of the 23 patients remaining in the study was 62.30  $\pm$  10.29 years, with a median duration of T2DM of 7 years (3-12 years). Despite the metabolic imbalance, only 8 patients (34.78%) reported a weight loss prior to initiating insulin therapy, with values ranging from 2 to 15 kg. The initial insulin dose was 18.96  $\pm$  6.32 units/day, and the final visit dose was 33.34  $\pm$  18.66 units/day. Regarding associate treatment, 21 patients (91.3%) received metformin, 16 patients (69.6%) were treated with angiotensin inhibitors, 1 patient (4.3%) with sartans, 8 (34.78%) with beta-blockers, 16 (69.6%) received statin therapy, and 5 patients (21.7%) had fenofibrates.

The evolution of clinical and paraclinical parameters is shown in Table 1. After the first year of insulin therapy the patients presented a significant improvement of parameters regarding glycemic control. HbA1c and basal glycaemia decreased progressively after first year of insulin therapy from 10.23 $\pm$ 1.94% to 7.63 $\pm$ 0.78% (p<0.001) and from 248.17 $\pm$ 61.84 mg/dl to 151.89 $\pm$ 41.72 mg/dl, respectively. Twenty patients (86.95%) presented weight gain, being observed a significant increase in body weight (p=0.005), with a median value of 2.4 kg (1.6; 5.5), but with no significant increase in body fat mass or percent of body fat. A significant increase of HDL-cholesterol and decrease of triglycerides was also recorded.

Adiponectin level was significantly reduced after one year comparative with initial level, while irisin presented significantly

Table 1. The evolution of clinical and paraclinical parameters during one year, after insulin therapy initiation

Parameter	Initial visit N=23	3 month visit N=23	6 month visit N=23	12 month visit N=23	P
HbA1c (%)	10.23±1.94	8.12±1.36	7.76±1.15	7.63±0.78	<0.001
BG (mg/dl)	248.17±61.84	157.97±61.36	167.84±39.39	151.89±41.72	<0.001
Weight (kg)	83.26±14.04	83.99±13.50	85.31±13.81	86.16±14.43	0.005
BMI (kg/m <sup>2</sup> )	29.8 (27.00;32.70)	30.4 (27.90;32.60)	30.9 (28.00;32.90)	31.6 (27.80;33.40)	0.01
PBF (%)	32.94±10.63	32.13±9.62	32.81±9.23	33.45±9.82	0.262
VFA (cm <sup>2</sup> )	137.66±39.87	141.23±36.79	138.53±32.62	141.91±37.37	0.608
SMM (kg)	31.00±7.15	31.66±7.11	31.85±7.51	31.97±7.54	0.005
BFM (kg)	28.3 (21.10;33.20)	27.2 (20.40;32.60)	27.2 (21.70;33.60)	27.5 (22.90;35.30)	0.128
SBP (mmHg)	142.91±20.74	145.21±16.02	138.65±16.43	142.60±16.51	0.434
DBP (mmHg)	82.17±9.43	82.65±9.87	83.34±10.07	81.69±10.87	0.904
TC (mg/dl)	196.87±69.86	170.84±44.12	176.14±43.37	184.22±49.77	0.16
LDL-cholesterol (mg/dl)	112.05±54.61	94.07±34.50	100.88±38.96	102.73±42.57	0.33
HDL-cholesterol (mg/dl)	39.03±12.59	44.55±15.37	43.17±12.12	44.18±13.02	0.012
Triglycerides (mg/dl)	180 (121;303)	159 (106;191)	146 (102;204)	145 (107;256)	0.014

N- Number of patients; BG – basal glycaemia; BMI – body mass index; PBF – percent body fat; VFA – visceral fat area; SMM – skeletal muscle mass; BFM – body fat mass; SBP – systolic blood pressure; DBP – diastolic blood pressure; TC – total cholesterol; LDL-cholesterol – low-density lipoprotein cholesterol; HDL-cholesterol – high-density lipoprotein cholesterol.

Table 2. Adiponectin, IL-1 $\beta$  and irisin level evolution after insulin therapy initiation

Parameter	Initial visit N=23	6 month visit N=23	12 month visit N=23	P
Adiponectin (mg/L)	7.06 (3.98;10.01)	7.44 (5.07;10.85)	4.81 (3.02;6.95)	0.044
IL-1 $\beta$ (pg/ml)	5.45 (3.54;7.22)	4.71 (2.65;6.33)	5.15 (3.65;6.70)	0.076
Irisin (ng/ml)	0.65 (0.00;1.63)	1.60 (0.99;1.84)	1.37 (0.57;1.70)	0.044

increased levels after the first year of insulin therapy, with no significant modification in IL-1 $\beta$  circulating levels (Table 2). Stratification for insulin dose showed a progressively significant reduction of adiponectin level after one year of insulin therapy in patients with more than 0.3 units of insulin/kg body weight/day ( $p=0.019$ ), but with no significant reduction in patients with less than 0.3 units of insulin/kg body weight/day.

The circulating levels of IL-1 $\beta$  presented significantly lower levels after one year of insulin therapy ( $p=0.038$ ) in patients which had lower initial levels of LDL-cholesterol. The other factors like age, gender, T2DM duration, initial BMI, body composition parameters, associated treatment or insulin dose had no significant influence on IL-1 $\beta$  levels.

Regarding the circulating levels of irisin, a significant increase was observed in patients treated with fenofibrate ( $p=0.001$ ). Other factors like age, gender, T2DM duration, initial BMI, body composition parameters, insulin dose, adiponectin and IL-1 $\beta$  levels were not influencing irisin levels.

## Discussions

Despite the anti-inflammatory effect of insulin, in patients with T2DM, insulin therapy appears to increase oxidative stress and inflammatory markers (Palem & Abraham 2015), with a direct correlation to insulin dose. In a recent published study, the level of high-sensitivity C-reactive protein was significantly increased in the third tertile of insulin dose/body weight, compared with the first tertile (Bala *et al* 2018). Accordingly, the interleukin-1 (IL-1) cytokine family is linked to inflammation-related diseases, including T2DM, while IL-1 $\beta$ , in particular, has been shown to be a target for reducing inflammation with potential therapeutic benefits (Donath *et al* 2009). This hypothesis was confirmed by the recently published results of the Phase III Cantos Study (The Canakinumab Anti-inflammatory Thrombosis Outcomes Study) involving 10,061 patients. Among patients, (approximately 40% with T2DM with a history of myocardial infarction and C-reactive protein  $\geq 2$  mg/dl), the anti-inflammatory therapy targeting IL-1 $\beta$  proved to reduce recurrent CV events compared to placebo, regardless the level of lipids (Ridker *et al* 2017).

Within our study, the variation of IL-1 $\beta$  during insulin treatment was insignificant, with no significant difference between the initial and the final values. Even though, initially, we observed a decrease in IL-1 $\beta$  levels, afterwards its level increased and the difference became insignificant after the first year of insulin therapy. We already know that acute administration of insulin leads to an initial decrease in inflammation markers (Dandona *et al* 2001). By reducing metabolic stress induced by hyperglycemia and direct suppression of proinflammatory cytokines, insulin potentially plays an anti-inflammatory role (Sun *et al* 2014). Considering these, although inflammation increases insulin resistance, there was no correlation between IL-1 $\beta$  level and total insulin dose in our study.

Adiponectin and inflammatory markers are related with insulin mediated turnover of the glucose (Nayak *et al* 2010). Adiponectin plays an important role in glucose metabolism by increasing muscle glucose uptake and insulin-sensitivity, by a direct vasodilatory effect on muscle microvasculature (Zhao *et al* 2014). In our study, a non-significant increase was observed after six months of insulin therapy, in contrast with Wang *et al* study, where a significant increase was observed in the same period of time (Wang *et al* 2015). In another study, a significant increase of adiponectin occurred in a cohort of 84 Chinese patients with T2DM after 3 months of insulin therapy, but no further increment was observed at 12 months (Zhang Q *et al* 2016). Up to a point, the same results were observed in both studies: an increase in adiponectin levels after 6 months of insulin therapy initiation, even though in our study the results were statistically insignificant, but in contrast in our study a significant decrease in the circulating level of adiponectin after the first year of insulin therapy was noticed. Further, the same process of adiponectin decrease was observed in a different study published by Fujita *et al* (2005). This phenomenon of insulin suppression on adiponectin was observed in mice, process explained by the hyperinsulinemic status caused by exogenous administration of insulin (Fujita H *et al* 2005). Similarly, in our study, the progressive and significant decrease of the adiponectin occurred in patients with a higher dose of insulin (more than 0.3 units of insulin/kg body weight/day). This phenomenon is explained by the fact that the low level of adiponectin decreases the oxidation of free fatty acids, and by this increasing their circulating level and insulin resistance. Consequently, higher doses are needed for glycemic control (Sheng & Yang 2008). Irisin is a myokine discovered in 2012 by Boström *et al* which has the ability to turn certain types of white tissue into brown tissue, potentially promoting weight loss by increasing thermogenesis, with a beneficial role in reducing insulin resistance (Boström *et al* 2012). The presence of irisin's precursors in adipose tissue suggests that it can also be an adipokine (Roca-Rivada *et al* 2013; Moreno-Navarrete *et al* 2013). Also, a recently published study on irisin highlighted that irisin can be an independent risk biomarker of CV disease in T2DM; the lower levels increasing by 1.6 fold the risk for CVD (El-Lebedy *et al* 2018). The physical activity, the high protein diet and some drugs like metformin, insulin, exenatide, simvastatin and fenofibrate treatment have a positive effect on irisin circulating levels, while the presences of obesity have a negative effect, lowering irisin levels (Mahgoub *et al* 2018).

Within our study, the circulating levels of irisin increased significantly after the first 6 months, afterwards starting to decrease. Its variation between the initial and final moment of insulin therapy remained significantly different. Also a significant increase in irisin levels was observed in patients treated with fenofibrate. This effect was also observed in mice study, and the explanation is that PPAR- $\alpha$  activation produced by fenofibrate stimulates formation of beige cells in subcutaneous white adipose tissue (Rachid *et al* 2015). On the other hand in patients with T2DM and hypertriglyceridemia, administration of fenofibrate decreases irisin levels, probably due to the reduction of irisin resistance (Feng *et al* 2015). Generally, irisin levels are lower in T2DM patients as compared with control group and no correlations with anthropometric or metabolic parameters were found (Liu *et al* 2013). These findings are supported by our study as well, since no significant clinical or metabolic parameter influencing the irisin level was found.

## Conclusions

The initiation of insulin therapy in people with T2DM is associated with a significant decrease in adiponectin and a significant increase in irisin after the first year of insulin therapy. The circulating IL-1 $\beta$  level was not significantly modified. For a better understanding of the role and influence of adiponectin, IL-1 $\beta$  and irisin in T2DM disease, further studies including a larger number of patients and a bigger follow up period are necessary. Also, more research studies should be focused on the potential use of irisin as an independent risk biomarker of CV disease in T2DM. The correlation of the adiponectin level with the dose of insulin can be explained by the fact that the low level of adiponectin decreases the oxidation of free fatty acids, and by this increasing their circulating level and insulin resistance. Considering this, the increase of insulin resistance can mask the anti-inflammatory effect of the insulin observed in other studies.

## Conflict-of-interest statement

Anca-Elena Crăciun declares speaker fees, sponsorships and consultancy fees from AstraZeneca, Sanofi, Eli Lilly, Servier, Merck Sharpe&Dhome, Mylan, Novo Nordisk, Amgen.

## References

- Aleidi S, Issa A, Bustanji H, Khalil M, Bustanji Y. Adiponectin serum levels correlate with insulin resistance in type 2 diabetic patients. *Saudi Pharm J.* 2015;23:250–6.
- Aljada A, Ghanim H, Saadeh R, Dandona P. Insulin inhibits NFkappaB and MCP-1 expression in human aortic endothelial cells. *J Clin Endocrinol Metab.* 2001;86:450–453.
- Asleh R, Sheikh-Ahmad M, Briasoulis A, Kushwaha SS. The influence of anti-hyperglycemic drug therapy on cardiovascular and heart failure outcomes in patients with type 2 diabetes mellitus. *Heart Fail Rev.* 2018;23(3):445-459.
- Bala C, Rusu A, Ciobanu DM, Craciun AE, Roman G. The association study of high-sensitivity C-reactive protein, pentraxin 3, nitrotyrosine, and insulin dose in patients with insulin-treated type 2 diabetes mellitus. *Ther Clin Risk Manag* 2018;14:955–963.

- Bonds DE, Miller ME, Bergenstal RM, Buse JB, Byington RP, Cutler JA et al. The association between symptomatic, severe hypoglycaemia and mortality in type 2 diabetes: retrospective epidemiological analysis of the ACCORD study. *BMJ*. 2010;340:b4909
- Boström P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC et al. A PGC 1-alpha-dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature*. 2012;481(7382):463–468.
- Dandona P, Chaudhuri A, Ghanim H, Mohanty P. Insulin as an anti-inflammatory and antiatherogenic modulator. *J Am Coll Cardiol*. 2009;53(Suppl.S):S14–20.
- Dandona P, Aljada A, Mohanty P, Ghanim H, Hamouda W, Assian E et al. Insulin Inhibits Intranuclear Nuclear Factor  $\kappa$ B and Stimulates I $\kappa$ B in Mononuclear Cells in Obese Subjects: Evidence for an Anti-inflammatory Effect? *J Clin Endocrinol Metab*. 2001;86(7):3257–65.
- Donath MY, Boni-Schnetzler M, Ellingsgaard H, Ehses JA. Islet inflammation impairs the pancreatic beta-cell in type 2 diabetes. *Physiology (Bethesda)* 2009; 24:325–331.
- El-Lebedy DH, Ibrahim AA, Ashmawy IO. Novel adipokines vaspin and irisin as risk biomarkers for cardiovascular diseases in type 2 diabetes mellitus. *Diabetes Metab Syndr*. 2018 Apr 12. pii: S1871-4021(18)30113-9
- Enerback S. Human brown adipose tissue. *Cell Metab*. 2010;11(4):248–252.
- Feng X, Gao X, Jia Y, Zhang H, Pan Q, Yao Z et al. PPAR-alpha agonist fenofibrate decreased serum irisin levels in Type 2 diabetes patients with hypertriglyceridemia. *PPAR. Res* 2015; 2015: 924131.
- Ferrannini E, DeFronzo RA. Impact of glucose-lowering drugs on cardiovascular disease in type 2 diabetes. *Eur Heart J*. 2015;36:2288–2296.
- Fujita H, Fujishima H, Koshimura J, Hosoba M, Yoshioka N, Shimotomai T et al. Effects of antidiabetic treatment with metformin and insulin on serum and adipose tissue adiponectin levels in db/db mice. *Endocr J*. 2005;52(4):427–33.
- Gamble JM, Simpson SH, Eurich DT, Majumdar SR, Johnson JA. Insulin use and increased risk of mortality in type 2 diabetes: a cohort study. *Diabetes Obes Metab*. 2010;12(1):47–53.
- Gerstein HC, Bosch J, Dagenais GR, Diaz R, Jung H, Maggioni AP et al (ORIGIN Trial Investigators). Basal insulin and cardiovascular and other outcomes in dysglycemia. *N Engl J Med*. 2012;367:319–328.
- Grover A, Padginton C, Wilson MF, Sung BH, Izzo JL, Dandona P. Insulin attenuates norepinephrine-induced venoconstriction. An ultrasonographic study. *Hypertension*. 1995;25:779–784.
- Herman ME, O'Keefe JH, Bell DSH, Schwartz SS. Insulin Therapy Increases Cardiovascular Risk in Type 2 Diabetes. *Prog Cardiovasc Dis*. 2017 Nov - Dec;60(3):422–434.
- Holman RR, Paul SK, Bethel MA, Matthews DR, Neil HA. 10-year follow-up of intensive glucose control in type 2 diabetes. *N Engl J Med* 2008; 359: 1577– 1589
- Huh JY, Panagiotou G, Mougios V, Brinkoetter M, Vamvini MT, Schneider BE et al. FNDC5 and irisin in humans: I. Predictors of circulating concentrations in serum and plasma and II. mRNA expression and circulating concentrations in response to weight loss and exercise. *Metabolism*. 2012; 61(12):1725–1738.
- Inbody 720 User's Manual. 1996-2004 Biospace Co, Ltd.; Available at: <http://www.bodyanalyse.no/docs/720%20users%20manual.pdf>. Accessed on 28th of December 2017
- Kadowaki T, Yamauchi T, Kubota N, Hara K, Ueki K, Tobe K. Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. *J Clin Invest*. 2006;116:1784–1792.
- Liu JJ, Wong MD, Toy WC, Tan CS, Liu S, Ng XW, et al. Lower circulating irisin is associated with type 2 diabetes mellitus. *J Diabetes Complications*. 2013 Jul-Aug;27(4):365–9.
- Mahgoub MO, D'Souza C, Al Darmaki RSMH, Baniyas MMYH, Adeghate E. An update on the role of irisin in the regulation of endocrine and metabolic functions. *Peptides*. 2018;104:15–23.
- Moreno-Navarrete JM, Ortega F, Serrano M, Guerra E, Pardo G, Tinahones F, et al. Irisin is expressed and produced by human muscle and adipose tissue in association with obesity and insulin resistance. *J. Clin. Endocrinol. Metab*. 2013;98:E769–778.
- Muis MJ, Bots ML, Grobbee DE, Stolk RP. Insulin treatment and cardiovascular disease; friend or foe? A point of view. *Diabet Med*. 2005;22(2):118–26.
- Nayak BS, Ramsingh D, Gooding S, Legall G, Bissram S, Mohammed A et al. Plasma adiponectin levels are related to obesity, inflammation, blood lipids and insulin in type 2 diabetic and non-diabetic Trinidadians. *Prim Care Diabet*. 2010;4:187–92.
- Nicholson T, Church C, Baker DJ, Jones SW. The role of adipokines in skeletal muscle inflammation and insulin sensitivity. *J Inflamm (Lond)*. 2018;15:9.
- Palem SP, Abraham PA. Study on the level of oxidative stress and inflammatory markers in type 2 diabetes mellitus patients with different treatment modalities. *J Clin Diagn Res*. 2015;9(9):BC04–7.
- Rachid TL, Penna-de-Carvalho A, Bringhenti I, Aguila MB, Mandarim-deLacerda CA, Souza-Mello V. Fenofibrate (PPARalpha agonist) induces beige cell formation in subcutaneous white adipose tissue from diet-induced obese mice. *Mol. Cell. Endocrinol*. 2015; 402: 86–94.
- Riddle MC, Ambrosius WT, Brillon DJ, Buse JB, Byington RP, Cohen RM et al. Epidemiologic relationships between A1C and all-cause mortality during a median 3.4-year follow-up of glycemic treatment in the ACCORD trial. *Diabetes Care*. 2010;33:983–990.
- Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C et al. Antiinflammatory Therapy with Canakinumab for Atherosclerotic Disease. *N Engl J Med*. 2017;377(12):1119–31.
- Roca-Rivada A, Castela C, Senin LL, Landrove MO, Baltar J, Belén Crujeiras A et al. FNDC5/irisin is not only a myokine but also an adipokine. *PLoS One* 8. 2013: e60563.
- Sheng T, Yang K. Adiponectin and its association with insulin resistance and type 2 diabetes. *J Genet Genomics*. 2008;35(6):321–6.
- Sun Q, Li J, Gao F. New insights into insulin: The anti-inflammatory effect and its clinical relevance. *World J Diabetes*. 2014; 5(2): 89–96.
- Timmons JA, Baar K, Davidsen PK, Atherton PJ. Is irisin a human exercise gene? *Nature*. 2012;488(7413):E9–E10.
- U.K. Prospective Diabetes Study (UKPDS) Group. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). *Lancet* 1998; 352: 837– 853
- Wang WD, Xing L, Teng JR, Li S, Mi NA. Effects of basal insulin application on serum visfatin and of adiponectin levels in type 2 diabetes. *Exp Ther Med*. 2015;9(6):2219–2224
- Yamamoto S, Matsushita Y, Nakagawa T, Hayashi T, Noda M, Mizoue T. Circulating adiponectin levels and risk of type 2 diabetes in the Japanese. *Nutr Diabetes*. 2014; 4(8): e130.
- Zhang Qa, Zhao X, Chen M, Fang Z, Chen Y, Wang Y. Weight gain and changes in plasma adiponectin and leptin concentrations after 12-month insulin intensive therapy for Chinese male patients with newly diagnosed type 2 diabetes. *Obes Res Clin Pract*. 2016;10(5):553–563.
- Zhao L, Fu Z, Liu Z. Adiponectin and Insulin Crosstalk: The Microvascular Connection. *Trends in cardiovascular medicine*. 2014;24(8):319–324.

**Authors:**

- Cristian-Ioan Crăciun, Department of Pharmacology, Toxicology and Clinical Pharmacology, University of Medicine and Pharmacy "Iuliu Hatieganu" Cluj-Napoca, 6 Louis Pasteur Street, Cluj-Napoca 400349, Romania, 400349; email: cristian.craciun@umfcluj.ro
- Anca-Elena Crăciun, Department of Diabetes, Nutrition and Metabolic Diseases, University of Medicine and Pharmacy "Iuliu Hatieganu" Cluj-Napoca; „Regina Maria” Clinic, Cluj-Napoca 24 Pasteur Street, Cluj-Napoca 400349, Romania; e-mail: anca.craciun@umfcluj.ro; doctor.craciun@yahoo.com
- Ștefan Cristian Vesa, Department of Pharmacology, Toxicology and Clinical Pharmacology, University of Medicine and Pharmacy "Iuliu Hatieganu" Cluj-Napoca, 6 Louis Pasteur Street, Cluj-Napoca 400349, Romania, 400349; email: stefanvesa@gmail.com
- Raluca Maria Pop, Department of Pharmacology, Toxicology and Clinical Pharmacology, University of Medicine and Pharmacy "Iuliu Hatieganu" Cluj-Napoca, 6 Louis Pasteur Street, Cluj-Napoca 400349, Romania, 400349; email: raluca\_parlog@yahoo.com
- Corina Ioana Bocșan, Department of Pharmacology, Toxicology and Clinical Pharmacology, University of Medicine and Pharmacy "Iuliu Hatieganu" Cluj-Napoca, 6 Louis Pasteur Street, Cluj-Napoca 400349, Romania, 400349; email: corina.bocsan@umfcluj.ro
- Anca Dana Buzoianu, Department of Pharmacology, Toxicology and Clinical Pharmacology, University of Medicine and Pharmacy "Iuliu Hatieganu" Cluj-Napoca, 6 Louis Pasteur Street, Cluj-Napoca 400349, Romania, 400349; email: abuzoianu@umfcluj.ro

**Citation** Crăciun CI, Crăciun AE, Vesa S, Pop RM, Bocșan CI, Buzoianu AD. The changes of inflammatory markers and irisine level in the first year of insulin therapy in type 2 diabetes patients. *HVM Bioflux* 2018;10(2):79-84.

**Editor** Antonia Macarie

**Received** 17 May 2018

**Accepted** 6 June 2018

**Published Online** 28 June 2018

**Funding** None reported

**Conflicts/  
Competing  
Interests** None reported